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                  U.S. patents
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                  CAS REGISTRY
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=> s (alpha-1-antitrypsin) (4A) (plasma or serum)
L1        3833 (ALPHA-1-ANTITRYPSIN) (4A) (PLASMA OR SERUM)

=> s L1 (6A) (human or sapiens)
L2         624 L1 (6A) (HUMAN OR SAPIENS)

=> s ((alpha-1-antitrypsin) or AAT or A1AT) (P) (recombinant or vector or plasmid
or transfection or coli or yeast)
L3        2492 ((ALPHA-1-ANTITRYPSIN) OR AAT OR A1AT) (P) (RECOMBINANT OR VECTO
               R OR PLASMID OR TRANSFECTION OR COLI OR YEAST)

=> s ((alpha-1-antitrypsin) or AAT or A1AT) (P) (glycosylation or deglycosylated or
endoglycosidase H)
L4         460 ((ALPHA-1-ANTITRYPSIN) OR AAT OR A1AT) (P) (GLYCOSYLATION OR
               DEGLYCOSYLATED OR ENDOGLYCOSIDASE H)

=> s l2 and l3 and l4
L5           5 L2 AND L3 AND L4

=> s l5  and (lyophilized or lyophilization or lyophilizing or (freeze dried) or
(speed vac) or (dried))
L6           0 L5 AND (LYOPHILIZED OR LYOPHILIZATION OR LYOPHILIZING OR (FREEZ
               E DRIED) OR (SPEED VAC) OR (DRIED))

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=> s l2 and l4 and (lyophilized or lyophilization or lyophilizing or (freeze dried)
or (speed vac) or (dried))
L7           0 L2 AND L4 AND (LYOPHILIZED OR LYOPHILIZATION OR LYOPHILIZING
               OR (FREEZE DRIED) OR (SPEED VAC) OR (DRIED))

=> s l2 and (lyophilized or lyophilization or lyophilizing or (freeze dried) or
(speed vac) or (dried))
L8           5 L2 AND (LYOPHILIZED OR LYOPHILIZATION OR LYOPHILIZING OR (FREEZ
               E DRIED) OR (SPEED VAC) OR (DRIED))

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L9           5 DUPLICATE REMOVE L8 (0 DUPLICATES REMOVED)

=> d l9 1-5 bib ab

L9   ANSWER 1 OF 5  CAPLUS  COPYRIGHT 2009 ACS on STN
AN   1990:473455  CAPLUS
DN   113:73455
OREF 113:12325a,12328a
TI   Production of alpha1-proteinase inhibitor (human)
AU   Hein, R. H.; Van Beveren, S. M.; Shearer, M. A.; Coan, M. H.; Brockway, W.
     J.
CS   Cutter Biol., Miles Inc., Berkeley, CA, USA
```

SO European Respiratory Journal (1990), 3(Suppl. 9), 16s-20s
CODEN: ERJOEI; ISSN: 0903-1936
DT Journal
LA English
AB A method for large scale isolation of α1-proteinase inhibitor (α1-PI) is described. This method employs waste Cohn fraction IV-1 as the starting material and involves fractional precipitation with polyethylene glycol followed by ion exchange chromatog. on DEAE-Sepharose. The process also incorporates a ten hour heat-treatment step at 60° to reduce or eliminate the risk of transmission of viral disease. The final product, having a purity of .apprx.60%, is freeze-dried . This preparation behaves almost identically to the α1-PI in plasma and is suitable for replacement therapy in hereditary emphysema.
OSC.G 2 THERE ARE 2 CAPLUS RECORDS THAT CITE THIS RECORD (2 CITINGS)

L9 ANSWER 2 OF 5 CAPLUS COPYRIGHT 2009 ACS on STN
AN 1989:219067 CAPLUS
DN 110:219067
OREF 110:36259a,36262a
TI Chromatogrpahic purification of .alpha.1-antitrypsin from human plasma cryopreceipitate fractions for medicaments
IN Burnouf, Thierry
PA Centre Regional de Transfusion Sanguine de Lille, Fr.
SO Fr. Demande, 8 pp.
CODEN: FRXXBL
DT Patent
LA French
FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	FR 2610633	A1	19880812	FR 1987-1403	19870205
	FR 2610633	B1	19920918		
	EP 282363	A2	19880914	EP 1988-400235	19880202
	EP 282363	A3	19881005		
	EP 282363	B1	19920909		
	R: AT, BE, CH, DE, ES, FR, GB, GR, IT, LI, LU, NL, SE				
	AT 80309	T	19920915	AT 1988-400235	19880202
	ES 2051871	T3	19940701	ES 1988-400235	19880202
	JP 01056699	A	19890303	JP 1988-26406	19880205
PRAI	FR 1987-1403	A	19870205		
	EP 1988-400235	A	19880202		

AB A concentrate of α1-antitrypsin (AAT) is prepared from human plasma by chromatog. of cryoppt. fractions A or A + I [Kistler and Nitschmann (1962)] to obtain an AAT solution of ≥80%. Human plasma from cryopptn. was precipitated with EtOH at 10% and pH 7.4 and the supernatant was precipitated with EtOH at 19%, pH 5.85, and 5°. EtOH was removed from the supernatant by diafiltration and the solution was diluted to .apprx.15 g protein/L and chromatographed on DEAE-Sepharose CL-6B Fast Flow equilibrated with 0.15M NaOAc pH 5.2-6. The AAT-rich fraction was adjusted to pH 6.5 with glycine, concentrated, dialyzed, and further purified on Sephacryl S-200. Viral inactivation was affected by heating to 60° for 10 h in the presence of sorbitol (65 weight%; stabilizer). After diafiltration to remove the sorbitol and adjusting the protein concentration to .apprx.25 g/L, the solution was placed in ampules and lyophilized. The AAT had trypsin and elastase inhibiting activities of native AAT.
OSC.G 5 THERE ARE 5 CAPLUS RECORDS THAT CITE THIS RECORD (5 CITINGS)
RE.CNT 2 THERE ARE 2 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

L9 ANSWER 3 OF 5 CAPLUS COPYRIGHT 2009 ACS on STN
AN 1988:607352 CAPLUS
DN 109:207352
OREF 109:34215a,34218a
TI Purification of alpha-1-proteinase inhibitor. Preparation and properties of a therapeutic concentrate
AU Coan, Michael H.
CS Cutter Biol., Miles Inc., Berkeley, CA, 94701, USA
SO American Journal of Medicine (1988), 84(6A), 32-6
CODEN: AJMEAZ; ISSN: 0002-9343
DT Journal
LA English
AB Human α1-proteinase inhibitor (α1-antitrypsin) (I) was prepared as a lyophilized concentrate and was tested clin. in humans with I deficiency. I protein was purified from blood plasma (Cohn fraction IV-1) by precipitation and ion-exchange chromatog. The resulting product behaved almost identically to I in plasma, showing that the process is gentle and nondenaturing. To lower the risk of transmission of disease, the product was heat treated. Although this resulted in some aggregation of protein, no new antigenic sites were created. Biol., immunol., and physiol. studies showed that I thus prepared behaves normally.

L9 ANSWER 4 OF 5 BIOSIS COPYRIGHT (c) 2009 The Thomson Corporation on STN
AN 1976:236089 BIOSIS
DN PREV197662066089; BA62:66089
TI HUMAN SKIN PROTEASES SEPARATION AND CHARACTERIZATION OF 2 ALKALINE PROTEASES 1 SPLITTING TRYPSIN AND THE OTHER CHYMOTRYPSIN SUBSTRATES.
AU FRAKI J E; HOPSU-HAVU V K
SO Archiv fuer Dermatologische Forschung, (1975) Vol. 253, No. 3, pp. 261-276.
CODEN: ADMFAU. ISSN: 0003-9187.
DT Article
FS BA
LA Unavailable
AB Two alkaline proteases, one splitting preferentially the substrates of chymotrypsin (N-acetyl-L-tyrosine ethyl ester, ATEE) and the other those of trypsin (N-α-benzoyl-L-arginine ethyl ester, BAEE), were separated and partially purified by chromatography from human skin extract made in a buffer containing 1.07 mol/l KCl. The proteins soluble in dilute buffer were removed by a prior extraction. The enzymes could be separated effectively only in the presence of KCl at a high concentration since large molecular size aggregates or polymers were formed in solutions of low ionic strength. In the presence of 2 mol/l KCl the molecular size of the BAEE-hydrolyzing enzyme was 120,000 and that of the ATEE-hydrolyzing enzyme 30,000. The ATEE-hydrolyzing enzyme was purified by Sephadex G-100 gel filtration and DEAE-cellulose chromatography about 250-fold. It also hydrolyzed esters of tryptophan and phenylalanine as well as casein with optimum pH 7.8-8.2. The enzyme was inhibited effectively by LBTI [trypsin inhibitor from lima bean, type II.L.], SBTI [lyophilized trypsin inhibitor from soybean, type Is] and partially by Trasylol, TPCK [L-1-tosylamide-2-phenyl-ethychloro-methylketone] and TLCK [N-α-p-toysl-L-lysine-chloro methylketone·HCl], but not by E-600 [diethyl-p-nitrophenyl phosphate] and SH-modifiers. The hydrolysis of ATEE was doubled in the presence of 1 mol/l KCl, NaCl, KBr or NaBr, but that of casein was inhibited to some extent. Human serum and .alpha.-1-antitrypsin inhibited this enzyme but not C.hivin.1-inactivator. α-2-Macroglobulin did not protect it from inhibition by SBTI. The BAEE-hydrolyzing enzyme was purified by Sephadex G-100 gel filtration and

hydroxylapatite chromatography about 30-fold. It also split other esters of substituted basic amino acids as well as BAPA [N-α-benzoyl-DL-arginine-p-nitroanilide·HCl] and histone proteins with optimum pH 7.5-8.2. It was inhibited by Trasylol and TLCK, but not by LBTI, SBTI, OMTI, [trypsin inhibitor from ovomucoid, type II] TPCK, E-600, SH-modifiers, human serum, C.hivin.1-inactivator or α-1-antitrypsin. Neither of these enzymes is exactly similar to any of the enzymes already separated from human tissues or fluids.

L9 ANSWER 5 OF 5 CAPLUS COPYRIGHT 2009 ACS on STN
AN 1971:417858 CAPLUS
DN 75:17858
OREF 75:2849a,2852a
TI Bacterial inactivation of human serum alpha-1 antitrypsin
AU Moskowitz, Roland W.; Heinrich, Gerhard
CS Sch. Med., Case West. Reserve Univ., Cleveland, OH, USA
SO Journal of Laboratory and Clinical Medicine (1971), 77(5), 777-85
CODEN: JLCMAK; ISSN: 0022-2143
DT Journal
LA English
AB The study demonstrates loss of human serum alpha-1 antitrypsin activity in the presence of cultures of certain gram-neg. bacterial organisms, as well as by exposure to lyophilized culture supernate prepared from Pseudomonas aeruginosa. Antitrypsin inactivation was seen to develop within 11 hr after inoculation of P. aeruginosa into broth. Upon incubation of lyophilized antitrypsin inactivator (Al) with antitrypsin at 37°, inactivation of antitrypsin increased as a function of time. Al was stable at 56° and at pH 5 through 8. Soybean trypsin inhibitor was not inactivated by 4-fold the amount of Al required to inactivate an equivalent number of moles of alpha-1 antitrypsin. Identical peaks were eluted with Sephadex G-75 column chromatog. when Al and antitrypsin were fractionated sep. or after prior preincubation, supporting an enzymic, rather than binding, action of Al on antitrypsin. Al may play a role in inflammatory mechanisms involving human serum alpha-1 antitrypsin.
OSC.G 1 THERE ARE 1 CAPLUS RECORDS THAT CITE THIS RECORD (1 CITINGS)

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=>
=> s l2 and (glycosylation or deglycosylated or endoglycosidase H)
L10 19 L2 AND (GLYCOSYLATION OR DEGLYCOSYLATED OR ENDOGLYCOSIDASE H)

=>
=> s l10 and (lyophilized or lyophilization or lyophilizing or (freeze dried) or (speed vac) or (dried))
L11 0 L10 AND (LYOPHILIZED OR LYOPHILIZATION OR LYOPHILIZING OR (FREEZ E DRIED) OR (SPEED VAC) OR (DRIED))